



Letter

Identification of broad neutralizing antibodies against Omicron subvariants from COVID-19 convalescents and vaccine recipients

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Dear Editor,

Omicron (B.1.1.529) was designated a variant of concern (VOC) on November 26, 2021 (Callaway, 2021), and its subvariants BA.1, BA.2, and BA.3 emerged and circulated almost simultaneously (Desingu et al., 2022). BA.2 was more efficient in transmission and quickly overtook BA.1 to become the variant most frequently detected worldwide (Yamasoba et al., 2022a). Compared to the prototype SARS-CoV-2 spike protein (S), the BA.1 and BA.2 spike proteins harbor more than 30 mutations, of which 21 are identical between the two subvariants, while the BA.3 spike differs from BA.1 and BA.2 by 3 mutations in the receptor binding domain (RBD) (Fig. 1A). More recently, BA.4 and BA.5 (hereafter BA.4/5) emerged, sharing the same spike sequence and containing four additional mutations, Del69–70, L452R, F486V, and R493Q, compared with BA.2. BA.4/5 were detected first in South Africa and evolved independently of BA.2; they have spread widely and replaced BA.2 as the predominant VOC (Gruell et al., 2022b; Tegally et al., 2022). In addition, BA.2.75, derived from the BA.2 subvariant, harbors nine additional mutations in the spike protein compared with BA.2 (Fig. 1A). BA.4/5 and BA.2.75 have led to the continuous emergence of novel Omicron subvariants, including BF.7 and BQ.1. These new subvariants may be driving waves of pandemics.

BA.1, BA.2, and BA.3 have proven to be considerably evasive of neutralization by sera from COVID-19 convalescents or vaccine

recipients and by monoclonal antibodies (mAbs) approved as therapeutics (Ai et al., 2022; Kurhade et al., 2022). Moreover, the BA.4/5 subvariants evade neutralization more efficiently than BA.2 (Cao et al., 2022; Hachmann et al., 2022). BA.2.75 has shown a lower capacity for escape from neutralization than BA.5 (Tan et al., 2022). Recent studies have highlighted that most therapeutic antibodies lose neutralizing activity completely or partially against Omicron subvariants (Gruell et al., 2022b; Yamasoba et al., 2022b). To date, only bebtelovimab has demonstrated potent neutralization activity against the Omicron BA.1, BA.2, BA.4/5, and BA.2.75 subvariants (Gruell et al., 2022b; Gruell et al., 2022a; Hentzien et al., 2022; Yamasoba et al., 2022a). Thus, identifying additional antibody candidates for neutralizing SARS-CoV-2 variants, especially emerging Omicron subvariants, is urgently needed. In this study, we tested the neutralizing profiles of a panel of mAbs previously isolated from COVID-19 convalescents and vaccine recipients and demonstrated five mAbs showing potent broad neutralizing activity against five major SARS-CoV-2 variants and Omicron subvariants.

Previously, we constructed a panel of mAbs from B cells of COVID-19 convalescents and vaccine recipients and demonstrated that 77 mAbs could neutralize pseudotyped prototype SARS-CoV-2 spike virus (Wuhan-Hu-1, GenBank no. YP_009724390.1; defined by $\geq 50\%$ inhibition observed at 1 $\mu\text{g}/\text{mL}$ mAb) (Fig. 1B). Whether these mAbs show broad neutralizing activity against SARS-CoV-2 variants, especially

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Omicron subvariants, has not been demonstrated. Here, we tested the neutralizing profile against SARS-CoV-2 VOCs by using pseudotyped virus. Our results showed that only a proportion of these mAbs were able to neutralize pseudotyped viruses of Omicron subvariants, with 27 for BA.1 (35.1%), 35 for BA.2 (45.5%), 22 for BA.3 (28.6%), 23 for BA.4/5 (29.9%), 32 for BA.2.75 (41.6%), 16 for BF.7 (20.8%), and 7 for BQ.1 (9.1%) (Fig. 1B and C). These data indicated that the emerging Omicron subvariants represented by BF.7 and BQ.1 had a greater ability to escape neutralization, especially BQ.1, which exhibited the largest neutralization resistance (Fig. 1C). All the mAbs bound to prototype S1, and 96.1% of the mAbs (74/77) bound to the RBD, of which 72.7% (56/77) of the mAbs showed competition for ACE2 binding (Fig. 1B). Of the Omicron-neutralizing mAbs, approximately two-thirds competed with ACE2, with 63.0% for BA.1 (17/27), 62.9% for BA.2 (22/35), 68.2% for BA.3 (15/22), 65.2% for BA.4/5 (15/23), 75.0% for BA.2.75 (24/32), 81.3% for BF.7 (13/16), and 57.1% for BQ.1 (4/7) (Fig. 1B). Collectively, these results suggest that most of these broadly neutralizing mAbs neutralized the prototype SARS-CoV-2 and Omicron subvariants by competitively binding to the RBD, thus interfering with the RBD-ACE2 interaction.

Next, we further characterized five mAbs that neutralized prototype SARS-CoV-2 by $\geq 95\%$ inhibition (observed at 1 $\mu\text{g}/\text{mL}$ mAb) and maintained potent neutralizing activity against Omicron subvariants BA.1 and BA.2 ($\geq 80\%$ inhibition at 1 $\mu\text{g}/\text{mL}$), which represent two distinct Omicron sublineages. The half-maximal inhibitory concentrations (IC_{50}) of these antibodies for prototype SARS-CoV-2 and Omicron subvariants were initially evaluated (Supplemental Table S1). These five antibodies showed potent neutralizing capacity in the majority of subvariants with a distinct neutralizing profile in the different subvariants (Fig. 1D–K). As expected, the BF.7 and BQ.1 subvariants exhibited greater neutralization resistance to these antibodies than the other subvariants. Compared to other antibodies, SCM15-45 was shown to be the most potent mAb for neutralizing prototype SARS-CoV-2, showing broad neutralizing profiles against Omicron subvariants (Fig. 1P). We further assessed the cross-binding reactivity and affinity of these five antibodies to different spike protein variants. All antibodies showed good reactivity and affinity for the spike proteins tested, with the exception of BA.1, which showed relatively weak activity (Supplementary Table S1). Among them, SCM11-12 bound to all tested spike proteins with a high activity, followed by SCM15-45. Sequence analysis revealed that these five mAbs originated from *IGHV3*, a V gene frequently used in the human antibody repertoire. The heavy chain CDR3 had a median length of 11 amino acids (aa) (ranging from 10 to 19 aa) and diverged from the germline sequence by 93.8%–98.0%. In addition, the light chain of these five mAbs was derived from *IGKV1*, *IGLV1* and *IGLV3*. The light chain CDR3 had a median length of 9 aa (ranging from 9 to 11 aa) and diverged from the germline sequence by 94.4%–98.3% (Supplementary Table S2). To test whether these five mAbs neutralized other VOCs, we performed neutralization assays for Alpha (B.1.1.7), Beta (B.1.351), Gamma (P.1), and Delta (B.1.617.2). All five mAbs efficiently neutralized these VOCs (Fig. 1L–O). SCM15-45 was the most potent mAb neutralizing these VOCs compared with the other mAbs (Fig. 1L–O). Together, we identified five mAbs that potently neutralized prototype SARS-CoV-2, five VOCs, and Omicron subvariants, with SCM15-45 showing a broad neutralizing profile.

The development and characterization of mAbs with broad neutralizing activity are vitally important for the prevention and therapy of infections with SARS-CoV-2 and its variants. Here, we identified a proportion of mAbs that had neutralizing activity against prototype SARS-CoV-2 and could also neutralize five major VOCs and Omicron subvariants recently shown to be prevalent worldwide. In addition, most of the mAbs we studied here lost neutralizing activity for Omicron subvariants to varying extents and exhibited distinct antibody escape patterns, such as in BF.7 and BQ.1, which are substantially more resistant, a finding consistent with serological results recently reported by other groups (Qu et al., 2022). The N460K and K444T mutations (present in BQ.1) and the R346T mutation (present in BF.7) are essential for the

enhanced resistance of the BQ.1 and BF.7 subvariants. The majority of the mAbs examined in this study showed good reactivity and affinity for the spike proteins, simultaneously showed binding to RBD and competition for ACE2 binding, which may explain that the potency in neutralizing the prototype and VOCs occurs primarily through interfering with RBD-ACE2 binding. Nevertheless, the mechanisms of neutralization require further investigation. A caveat of this study is that pseudotyped viruses were used to test for neutralization. Although previous studies have demonstrated a good correlation between pseudotype-based virus assays and assays using real viruses (Graham et al., 2021), the neutralizing profile of these mAbs needs to be further confirmed by authentic virus. The mAbs with broad neutralizing activity against SARS-CoV-2 and related variants developed here provide candidates for antibody therapy of COVID-19 and future vaccine design toward prophylactics of pancoronaviruses.

Footnotes

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